

Stuxnet detected, Pc breaks down

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Polycomb group (PcG) proteins were originally identified in *Drosophila*. They generally maintain gene silencing by forming multimeric complexes. Two main complexes, namely Polycomb repressive complex 2 (PRC2) and PRC1, have been described. PRC2 methylates histone H3 on lysine 27 (H3K27). PRC1, mainly composed of Polycomb (Pc), Polyhomeotic (Ph), Posterior sex combs (Psc) and dRing/Sce, has been shown to directly compact chromatin *in vitro*. The mutation or deficiency of these members causes posterior transformation of the body plan as homeotic (HOX) genes are depressed. In the past two decades, main advances have been achieved in the diversity of PRC compositions, the dynamic chromatin targeting mechanisms, the silencing mechanisms involving both histone modifications and chromatin compaction (Blackledge et al., 2015; Simon and Kingston, 2013). Meanwhile, more and more studies have been focusing on the upstream regulatory mechanisms of PcG expression. For example, the expression of CBX proteins, the orthologs of Pc in mammalian cells have been shown to be regulated at the transcriptional and post-transcriptional levels (briefly reviewed in (Gil and O’Loghlen, 2014)). Furthermore, PcG protein stability as well as their localization or enzymatic activities has also been reported to be regulated by post-translational modifications (PTMs), such as phosphorylation and ubiquitination (Niessen et al., 2009). In a recent issue of *Developmental Cell*, Du et al. described an additional layer of PTM-independent regulation on PcG (Pc) stability (Du et al., 2016) (Figure 1).

Taking advantage of an engineered insertion of anten-

na-specific Gal4 driver system to induce transcription, Du et al. performed a Gene Search-based mis-expression screen for developmental regulators located on the X chromosome in *Drosophila*. They identified a gene *Stx*, whose induction led to defective antenna development. Then the authors validated the phenotype by generating transgenic *flies* overexpressing *Stx* that displayed wing-to-haltere as well as antenna-to-leg transformation. Accordingly, high levels of *Stx* resulted in the de-repression of homeotic genes *Antp* and *Ubx*, etc. Therefore, overactive *Stx* has a general role in homeotic transformation, which is reminiscent of the loss of PcG functions. In contrast, the loss of *Stx* caused early lethality at the pupal stage of development, which could be rescued by deletion of one allele of *Pc*. Moreover, the authors showed that *Stx* and *Pc* display a reciprocal expression pattern. These findings strongly indicated an inverse functional relationship between *Stx* and *Pc*.

In addition to the genetic interaction, the physical interaction between *Stx* and *Pc* was confirmed by *in vivo* co-immunoprecipitation and *in vitro* GST-pull down assays. And a Pc-binding domain (PcB) was mapped in *Stx*. Interestingly, increased expression of wild type *Stx* but not the *Stx*ΔPcB mutant reduced the *Pc* expression, suggesting that the interaction is essential for *Stx* to downregulate the *Pc* protein level. The authors then wondered whether *Stx* directly affects *Pc* protein stability. Indeed, *Pc* protein is subject to proteasome degradation. Overexpressing *Stx* facilitates *Pc* degradation which could be reversed by the treatment of the proteasome inhibitors. On the contrary, depletion of *Stx* stabilizes *Pc* proteins. Based on these evidences, *Pc* breaks down at the high level of *Stx*. Hence the authors name the gene as *Stuxnet*, originally referring to a PC com-

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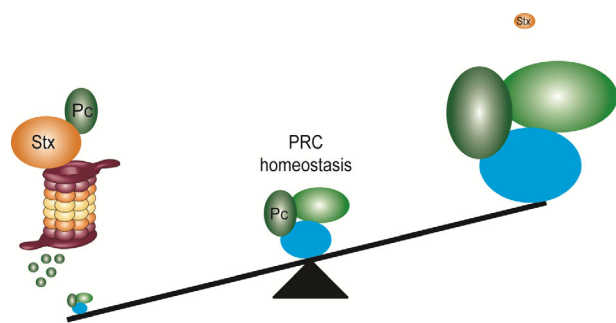


Figure 1 PRCs are dynamically regulated during development. Du et al. in a recent issue of *Developmental Cell* identified a new regulator Stx that controls PRC homeostasis by adapting Pc for proteasome degradation.

puter virus.

Protein ubiquitination is one of the major routes for proteasome degradation. However, to our surprise, the authors found that Pc degradation is not dependent on the lysine ubiquitination despite its existence on Pc. Then what is the mechanism? After a careful analysis of the known motifs, a Ubiquitin-like (UBL) domain in Stx intrigued the researchers. Similar to other UBL-domain containing proteins, Stx interacts with the 19S proteasome regulatory subunit Rpn10 in the nucleus dependent on the UBL domain. As the proteasome-interacting motif in the UBL domain forms an interacting surface with the 26S proteasome, Stx relays Pc to the degradation machinery. Moreover, the ubiquitination-independent proteasome degradation of Pc was phenocopied by the *in vivo* genetic models in which the Stx Δ UBL mutant stabilized Pc protein and therefore acted in a dominant negative fashion.

Furthermore, the authors convincingly demonstrated that Stx has a negative impact of Pc functions on chromatin *in vivo*. As shown by both imaging and chromatin immunoprecipitation (ChIP) assays, the general decreased binding of Pc on polytene chromosomes and the engineered Polycomb Response Element (PRE) was observed upon overexpression of full length Stx, rather than the Stx Δ PcB mutant. This is unlikely due to the decreased H3K27me3 levels as Stx does not seem to interact with and target PRC2 members.

Finally the authors demonstrated that this regulatory

mode is conserved in mammals. For yet unknown reasons, the mouse homolog of Stx (mStx) specifically targets Cbx4, one of the five Pc orthologs when co-expressing in 293 cells. But it remains a mystery whether mStx targets other Cbx proteins in distinct contexts. No matter Cbx4-specific targeting or not, the physiological and pathological significances of this regulatory mechanism await further specification in mammals.

PcG proteins have attracted attention of more than the development biologists in the past decades, because of their documented role in regulating cellular identity. This study, combining genetic manipulations in *Drosophila* and biochemical approaches, has clearly illustrated a previously unrecognized role of developmental regulators in modulating PcG stability. It opened a new avenue to understand the upstream regulatory mechanisms on the precise control of PcG expression and activity. In the future experiments, it will be worthwhile to sort out the signalings that affect Stx activity and bridge Stx-Pc interaction in physiological contexts. Interestingly, the mutations or aberrant expression of Stx has been found in cancers. Stx mutations were also identified in genetic defects such as autism spectrum disorders (ASDs). Though Stx may possess Pc-independent functions, it is tempting to speculate that the PcG activity is dysregulated due to the malfunction of Stx in these disease states.

Compliance and ethics The author(s) declare that they have no conflict of interest.

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